Natural Products

Total Synthesis of Apoptolidinone**

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Dedicated to Professor Amos B. Smith, III on the occasion of his 60th birthday.

Polyketide-derived secondary metabolites have long served as a source of structurally diverse and biologically active natural products.^[1] In the course of screening soil microorganisms for cell-specific apoptosis-inducing agents, Hayakawa and co-workers isolated the polyketide natural product apoptolidin (2) from *Nocardiopsis* sp.^[2] Apoptolidin induces

programmed cell death in E1A-transformed cells but not in normal cells.^[2a] Khosla, Salomon, and co-workers later correlated this cell-specific activity to the inhibition of mitochondrial F₀F₁-ATPase by apoptolidin as well as other polyketide natural products.^[3] Structurally, apoptolidin features an unsaturated 20-membered macrolide, a six-membered hemiketal, and three hexose sugars.^[2b] In 2001 Koert and co-workers reported the synthesis of apoptolidinone (1) and later the same year Nicolaou and co-workers described the total synthesis of apoptolidin.^[4-6] The latter total synthesis and other recent work has demonstrated that apoptolidin is a rather labile compound that undergoes a base-induced

acyl migration from the C19 to C20 hydroxy group to produce isoapoptolidin, [7] a compound that is less active against

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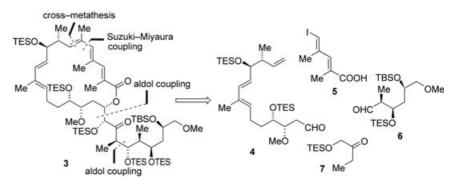
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mitochondrial F_0F_1 -ATPase. [7a] Although evaluation of the cytotoxicity of select analogues has suggested the three hexose sugars of apoptolidin to contribute significantly to the overall cytotoxicity of $\mathbf{2}$, [3c,5e,8] biological evaluation of the aglycone, apoptolidinone (1), has not been reported. We describe herein the total synthesis of apoptolidinone.

Besides the synthetic approach to apoptolidin (2) described by Toshima and co-workers, [6a] all previous synthetic strategies directed towards apoptolidin have relied on a linear approach in which the seco acid was assembled and subsequently subjected to a macrolactonization. To develop a more convergent assembly we retrosynthetically divided apoptolidinone into four fragments (4–7, Scheme 1). We planned to couple the four fragments through a combination of two diastereoselective aldol reactions, a Grubbs crossmetathesis reaction, and an intramolecular Suzuki–Miyaura cross-coupling reaction.

Construction of fragment **4** started from (*S*)-malic acid (**8**), which was converted into 3-methoxy-γ-butyrolactone (**9**) through a known four-step reaction sequence. Reduction of lactone **9** with DIBAL-H afforded lactol **10**, which on condensation with 1,3-propanedithiol afforded dithiane **11**. Swern oxidation of primary alcohol **11** provided aldehyde **12** in near quantitative yield. A five-carbon unit was introduced to aldehyde **12** by chelation-controlled addition of the Grignard reagent derived from bromide **13** to provide secondary alcohol **14** as a single isomer (Scheme 2). Bromide **13** was prepared from dihydrofuran according to the Kocien-



Scheme 1. Retrosynthetic analysis of apoptolidinone (1). TES = triethylsilyl, TBS = *tert*-butyldimethylsilyl.

ski procedure described by Koert and co-workers in their reported synthesis of apoptolidinone. [4a,10] A solution of **14** in dichloromethane was treated sequentially with iodine, trie-thylchlorosilane, and imidazole in one pot to provide vinyl iodide **16** in 90% overall yield from **14**. The dithiane group of **16** was hydrolyzed efficiently by using the Fetizon–Jurion procedure to provide aldehyde **17** in 68% yield (as well as recovered **16** (19%)). [11]

Homoallylic silyl ether **18** was produced by the asymmetric addition of the diisopropyl tartrate ester derived (*Z*)-crotylboronate reagent developed by Roush et al.^[12] to the pinacol ester of 3-boronoacrolein,^[13] followed by silylation (TESOTf, 2,6-lutidine) of the crude crotylation product. The *syn* homoallylic ether **18** (single diastereomer, 80 % *ee*) was

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Zuschriften

Scheme 2. Synthesis of fragment **4.** a) DIBAL-H, THF, $-78\,^{\circ}$ C; b) 1,3-propanedithiol, BF₃·OEt₂, CH₂Cl₂, 28 $\,^{\circ}$ C, 60% from **9**; c) (COCl)₂, DMSO, iPr₂NEt, $-78\,^{\circ}$ O°C, 98%; d) **13**, Mg, 1,2-dibromoethane, Et₂O, $-78\,^{\circ}$ C, 68%; e) I₂, CH₂Cl₂, 0 $\,^{\circ}$ C; f) TESCl, ImH, CH₂Cl₂, 28 $\,^{\circ}$ C, 90% from **14**; g) MeI (excess), K₂CO₃, MeCN/pH 7 buffer (4:1), 28 $\,^{\circ}$ C, 68% (plus 19% recovered **16**); h) **18**, [Pd(Ph₃P)₄], TlOH, THF/H₂O (3:1), 28 $\,^{\circ}$ C, 70%. DIBAL-H = diisobutylaluminum hydride, DMSO = dimethyl sulfoxide, ImH = imidazole.

obtained in 43 % yield over two steps. Suzuki–Miyaura cross-coupling between vinyl iodide **17** and vinyl boronate **18** provided diene **4** in 70 % yield (Scheme 2).^[14]

The stereochemical relationship between C19 and C20 (see 20) was established by a diastereoselective Mukaiyama aldol condensation between aldehyde 4 and enol silane (Z)-19 (derived from 1,2-butanediol in three steps) to afford ketone 20 as the major product with a 4:1 ratio of isomers (Scheme 3). The assigned C19-C20 relative stereochemistry rested on the observed coupling constant of the aldol product $(J_{19,20} = 3.5 \text{ Hz})$ and the 1,3-asymmetric induction model proposed by Evans and co-workers for β-methoxyaldehydes.^[15] Yamaguchi esterification of 20 with carboxylic acid 5 led to isolation of dienoate 21 in 83% yield. [16] Kinetic deprotonation (LHMDS, HMPA, THF, -78°C) of 21 followed by aldol condensation with aldehyde $6^{[17]}$ afforded syn aldol product 22 as a single isomer in 41 % yield (plus 30 % recovered 21).[18] After silvlation of 22 to give 23 we examined a series of cross-metathesis reactions with propenyl boronate to provide vinyl boronate 25.[19] The Grubbs second-generation catalyst 24 provided 25 in up to 30% yield (plus 30% recovered alkene 23). Remarkably, this reaction provided 25 as a single isomer that was immediately subject to an intramolecular Suzuki-Miyaura cross-coupling to give macrolactone 3 in 47-60% yield. Exhaustive desilylation of 3 provided apoptolidinone (1) in 61% yield. Our synthetic apoptolidinone matched, in all respects, the spectral data reported by Koert and co-workers for their synthetic sample.[4a]

Scheme 3. Synthesis of apoptolidinone (1). a) BF₃·OEt₂, CaH₂, CH₂Cl₂, -94°C, 50% (plus 34% recovered 4); b) 5, 2,4,6-trichlorobenzoyl chloride, Et₃N, DMAP, toluene, $-78 \rightarrow 28$ °C, 83%; c) LHMDS, THF, HMPA, -78°C, 2 h; then 6, THF, 41% (plus 30% recovered 21); d) TESOTf, 2,6-lutidine, CH₂Cl₂, 0°C, 81%; e) 24, isopropenyl pinacol boronic ester (18 equiv), CH₂Cl₂, reflux, 6 h, 30% (plus 30% recovered 23); f) [Pd(Ph₃P)₄], Tl(OEt), THF/H₂O (3:1), 28°C, 30 min, 60%; g) HF·py, THF, -10°C, 12 h; then -5°C, 5 h, 61%. DMAP=4-(dimethylamino)pyridine, LHMDS=lithium hexamethyldisilazide, HMPA=hexamethylphosphoramide, Tf=trifluoromethanesulfonyl.

In summary, apoptolidinone was synthesized from 3methoxy-γ-butyrolactone (9) in 14 steps (longest linear sequence). Key steps of the synthesis include two diastereoselective aldol reactions, a cross-metathesis reaction, and two Suzuki-Miyaura cross-coupling reactions. We anticipate that this synthesis will provide access to modified derivatives of apoptolidin for utilization in studies on the cell-specific cytotoxicity of the parent natural product. [20]

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- [20] We employed the synthetic route described herein to produce 2-4 mg of apoptolidinone, which is currently undergoing biological evaluation in a side-by-side comparison with apoptolidin itself.

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